



Available online at www.sciencedirect.com



Neuromuscular Disorders 23 (2013) 969-974



Polymorphism in the *TOMM40* gene modifies the risk of developing sporadic inclusion body myositis and the age of onset of symptoms

F.L. Mastaglia^{a,b,*}, A. Rojana-udomsart^a, I. James^b, M. Needham^{a,b}, T.J. Day^c, L. Kiers^c, J.A. Corbett^d, A.M. Saunders^e, M.W. Lutz^e, A.D. Roses^{e,f}, for the Alzheimer's Disease Neuroimaging Initiative¹

> ^a Australian Neuro-Muscular Research Institute, Centre for Neuromuscular and Neurological Disorders, The University of Western Australia, Queen Elizabeth II Medical Centre, Perth, Western Australia, Australia
> ^b Institute for Immunology & Infectious Diseases, Murdoch University, Western Australia, Australia
> ^c Departments of Neurology and Neurophysiology, Royal Melbourne Hospital & Department of Medicine, University of Melbourne, Parkville, Victoria, Australia
> ^d Department of Neurology, Concord Hospital, Concord, NSW, Australia
> ^c Duke University, Durham, NC 27705, USA

^fZinfandel Pharmaceuticals, Durham, NC 27705, USA

Received 25 June 2013; received in revised form 28 August 2013; accepted 10 September 2013

Abstract

A polyT repeat in an intronic polymorphism (rs10524523) in the *TOMM40* gene, which encodes an outer mitochondrial membrane translocase involved in the transport of amyloid- β and other proteins into mitochondria, has been implicated in Alzheimer's disease and *APOE-TOMM40* genotypes have been shown to modify disease risk and age at onset of symptoms. Because of the similarities between Alzheimer's disease and sporadic inclusion body myositis (s-IBM), and the importance of amyloid- β and mitochondrial changes in s-IBM, we investigated whether variation in poly-T repeat lengths in rs10524523 also influence susceptibility and age at onset in a cohort of 90 Caucasian s-IBM patients (55 males; age 69.1 ± 9.6). In carriers of *APOE* ϵ_3/ϵ_3 or ϵ_3/ϵ_4 , genotypes with a very long (VL) poly-T repeat were under-represented in s-IBM compared to controls and were associated with a later age at symptom onset, suggesting that these genotypes may be protective. Our study is the first to suggest that polymorphisms in genes controlling mitochondrial function can influence susceptibility to s-IBM and have disease modifying effects. However, further studies in other s-IBM populations are needed to confirm these findings, as well as expression studies of different *TOMM40* alleles in muscle tissue. © 2013 Elsevier B.V. All rights reserved.

Keywords: Sporadic IBM; TOMM40; APOE; Susceptibility; Age of onset

* Corresponding author at: Australian Neuro-Muscular Research Institute, Centre for Neuromuscular and Neurological Disorders, The University of Western Australia, Queen Elizabeth II Medical Centre, Perth, Western Australia, Australia. Tel.: +61 893461611; fax: +61 893463487.

E-mail address: francis.mastaglia@anri.uwa.edu.au (F.L. Mastaglia).

0960-8966/\$ - see front matter © 2013 Elsevier B.V. All rights reserved. http://dx.doi.org/10.1016/j.nmd.2013.09.008

1. Introduction

Sporadic inclusion body myositis (s-IBM) is a progressive degenerative and inflammatory myopathy which has a number of features in common with Alzheimer's disease (AD), such as a late onset (most commonly over the age of 50 years), abnormal accumulation of amyloid- β and other misfolded proteins,

¹ Data used in preparation of this article were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (adni. loni.ucla.edu). As such, the investigators within the ADNI contributed to the design and implementation of ADNI and/or provided data but did not participate in analysis or writing of this report. A complete listing of ADNI investigators can be found at: http://adni.loni.ucla.edu/wp-content/uploads/how_to_apply/ADNI_Acknowledgement_List.pdf.

mitochondrial dysfunction and oxidative stress [1–4]. However, studies to date have shown differences in genetic susceptibility for the two diseases. Whereas in AD apolipoprotein *APOE* ε 4 is a strong risk factor [5], in s-IBM there is no recognised association with *APOE* genotype [6,7], although APOE has been shown to co-localise with β -amyloid in vacuolated muscle fibres [8]. The strongest genetic association in s-IBM is with the *HLA-DRB1* and secondary *HLA-DRB* loci in the central MHC region [9–11]. In addition, in a phylogenetic analysis of mtDNA variants we demonstrated an association with the 4336G and 4580A D-loop variants in s-IBM, but not in AD [12].

The 'Translocase of Outer Mitochondrial Membrane 40' homologue (TOMM40) gene is adjacent to and in linkage disequilibrium with the APOE locus on chromosome 19. TOMM40 encodes the mitochondrial pore protein Tom40, which is part of the TOM complex and is involved in the passage of peptides and importation of amyloid- β into mitochondria [13]. Roses et al [14] first reported that in late-onset Alzheimer's disease (LOAD), carriers of the APOE $\varepsilon 3/\varepsilon 4$ genotype, with a VL-[very long] poly-T repeat from the ɛ3 strand and a L [Long] poly-T repeat from the ɛ4 strand at rs10524523 in intron 6 of TOMM40 had a higher disease risk and an earlier age at onset than individuals with the S [Short] poly-T on the APOE-TOMM40 linkage disequilibrium region. An association between carriage of the VL allele per se and disease risk was subsequently also found in other AD populations [15], but in the opposite direction, with carriage of the VL allele being found to be protective [16]. Subsequent observations demonstrated that the genotypes of both inherited alleles are in fact important in determining the age-at-onset, with homozygotes of the VL allele having the oldest age-at-onset curve and other genotypes being associated with an earlier onset [17].

In this study we investigated whether genetic variation in *APOE-TOMM40* also influences disease susceptibility and the age of symptom onset in s-IBM. Our hypothesis was that genetic variants of *TOMM40* may have differential effects on mitochondrial function in muscle that may impact on the risk of developing s-IBM and the tempo of the disease, and that some alleles of *TOMM40* may have a protective effect.

2. Materials and methods

2.1. Subjects

DNA was collected from 90 Caucasian s-IBM patients (55 males; age 69.1 ± 9.6) recruited at the Australian Neuromuscular Research Institute in Perth, the Royal Melbourne Hospital and Monash Medical Centre in Melbourne, and the Concord Repatriation and Royal North Shore Hospitals in Sydney, who fulfilled the

diagnostic criteria for definite or probable s-IBM [2,18]. All patients had a detailed clinical history taken, including the age-at-onset of the initial symptoms of limb muscle weakness, and a full neurological examination with grading of muscle strength on an expanded (10-point) Medical Research Council scale. The age-at-onset was determined from the recollection of the patients, and when available their spouses, of the year in which they first became aware of symptoms of lower limb or hand weakness, and ranged from 37 to 83 years (mean 60.4 ± 9.7 years). The mean disease duration at the time of DNA collection was 8.7 years. Muscle biopsy reports, and when necessary the biopsy slides, were reviewed. The study was approved by the Sir Charles Gairdner Hospital Human Ethics Committee (Approval Number 2006-073).

The control group comprised 205 individuals (mean age 76.0 ± 5.2 years) from the Alzheimer's Disease Neuroimaging Initiative (ADNI) Database. The ADNI was launched in 2003 by the National Institute on Aging. the National Institute of Biomedical Imaging and Bioengineering, the Food and Drug Administration and private pharmaceutical companies. The primary goal of ADNI has been to test whether serial magnetic resonance imaging, positron emission tomography, other biological markers, and clinical and neuropsychological assessment can be combined to measure the progression of mild cognitive impairment and early Alzheimer's disease. The Principal Investigator is Michael W. Weiner, MD, VA Medical Center and University of California - San Francisco. ADNI is the result of efforts of many co-investigators from a broad range of academic institutions and private corporations, and subjects have been recruited from over 50 sites across the U.S. and Canada.

2.2. Genotyping

DNA samples were plated on 96-well plates for longrange PCR and sequencing which was performed at Polymorphic DNA Technologies (Alameda, CA, USA), as described previously by Roses et al. [14] and Linnertz et al. [19]. Based on the length of the PCR product, alleles of rs10524523 were classified using the convention established by Roses et al for determining alleles: Short (S), \leq 19; Long (L) 20–29; Very Long (VL) \geq 30 [14,19]. *APOE* genotyping was performed as described previously [19].

2.3. Statistical methods

Frequencies were compared by chi-square or stratified Mantel-Haenszel tests as appropriate. Distributions of ages at onset adjusting for gender were compared via Cox proportional hazards models. Analyses were carried out using the TIBCO Spotfire S+ package ver 8.2 (TIBCO Software Inc., Palo Alto, California).

3. Results

b

0

0

15

20

3.1. APOE genotypes

APOE ε 3. Of these, 50 were homozygous, 26 carried ε 3/ ε 4, and nine carried $\varepsilon 2/\varepsilon 3$. The remaining five s-IBM cases carried $\varepsilon 2/\varepsilon 4$ (2), $\varepsilon 4/\varepsilon 4$ (2) or $\varepsilon 2/\varepsilon 2$ (1).

In keeping with the findings of Roses et al. [14], the distribution of poly-T repeat lengths among those homozygous for APOE ε 3 was strongly bimodal, with a gap between 17 and 25 repeats (Fig. 1). Analyses were confined to carriers of $\varepsilon 3/\varepsilon 3$ and $\varepsilon 3/\varepsilon 4$, within which carriage of VL was likely associated with APOE E3. Numbers of cases and controls carrying the VL repeat length according to the two APOE genotype groups are shown in Table 1. The odds ratios within the two groups are not significantly different (p = 0.44), while the combined Mantel-Haenszel odds ratio estimate of 0.47 is significantly less than one (p = 0.019, 95% CI 0.25–0.88). Carriage of a VL poly-T repeat length within these groups is thus significantly associated with protection.

3.3. Poly-T repeat length and AAO of s-IBM

The boxplots in Fig. 2 show the ages at onset for the four subgroups of s-IBM patients defined as $\varepsilon_3/\varepsilon_4$ or $\varepsilon_3/\varepsilon_4$ ε 3 carrying or not carrying VL. Note that there are only 7 individuals in the group $\varepsilon 3/\varepsilon 4$ with VL and these display more variability than the remaining subgroups. Kaplan-Meier plots demonstrating the age at onset distributions for individuals carrying and not carrying a VL poly-T length are given in Fig. 3. Whilst gender was not significant (p = 0.2), after adjusting for gender those with VL had a later age at onset overall (p = 0.038).

The majority (85/90) of s-IBM patients were carriers of

3.2. Poly-T repeat lengths in s-IBM and controls

4. Discussion

This is the first study to investigate the influence of polymorphism in the TOMM40 gene in s-IBM. Our findings show that among carriers of APOE ε 3 there is a significant association between carriage of rs10524523 genotypes including a very long (VL) poly-T repeat length allele and a reduced risk of s-IBM as in the case of AD [16]. Moreover, carriage of APOE-TOMM40 genotypes with a VL allele was associated with a later age at onset of symptoms. These findings therefore both point to the possibility that these genotypes have a protective effect and warrant further investigation.

Mitochondrial abnormalities are an important part of the pathological phenotype of s-IBM, and include raggedred and cytochrome oxidase (COX) deficient muscle fibres, ultrastructural abnormalities, and multiple somatic mtDNA deletions which are associated with defective synthesis of COX and other components of the respiratory enzyme chain [20-22]. Mitochondrial dysfunction and oxidative stress, which is increased in s-IBM muscle fibres [23–25], are known to stimulate aggregation of amyloidogenic proteins [26]. Moreover, over-expression of β APP and accumulation of amyloid- β are early changes in muscle fibres prior to the development of structural abnormalities [27], and over-expression of β APP in human myoblasts in vitro leads to the development of structural mitochondrial abnormalities and loss of COX activity [28]. Increased transport of amyloid-ß into mitochondria, or arrest of BAPP in the mitochondrial import pores, as has been demonstrated in AD [29,30], could interfere with COX activity and lead to increased generation of reactive oxygen species (ROS) [31]. Genetic variants of TOM40 could be associated with altered mitochondrial pore function and transport of $\beta APP/amyloid-\beta$ and other proteins into mitochondria. This could in turn lead to changes in energy metabolism and increased generation of ROS, which could contribute to impaired mitochondrial integrity and muscle fibre degeneration.

The rs10524523 locus may influence susceptibility to AD and s-IBM by modulating expression levels of TOMM 40 or APOE. Expression studies of TOMM 40 alleles with different poly-T repeat lengths in brain have produced varying findings. The study by Cruchaga et al. [16] on a small number of AD brain samples failed to show any differences in TOM40 cDNA levels with different rs10524523 alleles. However, in a more detailed analysis of a larger number of samples, Linnertz and colleagues have shown that the VL allele is associated with higher expression levels of TOMM40 and APOE mRNA, both in normal and AD brain specimens (unpublished). Similar studies have yet to be performed on muscle samples from s-IBM and normal subjects and will be important in determining if expression levels in muscle are also altered by carriage of VL allele bearing genotypes.

The present findings support our previous suggestion that multiple genes, both immune and non-immune, may



25

30

35



Table 1 Frequency of carriage of VL poly-T repeat length in $\epsilon 3/\epsilon 3$ and $\epsilon 3/\epsilon 4$ individuals.						
	ε3/ε3					
		NT 17				

	63/63		85/84	
	VL carriage	No VL carriage	VL carriage	No VL carriage
sIBM:	34	16	7	19
ADNI:	95	25	24	22
Odds ratio:	0.56			0.34

2/ 4

Mantel-Haenszel combined OR = 0.47 (p = 0.019).



Fig. 2. Box-plots of the ages at disease onset in subgroups of ϵ_3/ϵ_4 or ϵ_3/ϵ_3 s-IBM carriers, with or without VL alleles, showing a later age at onset in ϵ_3/ϵ_3 homozygotes with a VL allele.



Fig. 3. Kaplan-Meier plots showing a significantly later age at disease onset in individuals with genotypes carrying a VL allele.

contribute to susceptibility to s-IBM [3]. However, the findings need to be confirmed in other s-IBM patient cohorts and should also be investigated in patients of other genetic and racial backgrounds. In particular, the

association with age at onset requires further confirmation, in view of the potential limitation of self-reported data on the age of symptom onset in s-IBM. Identification of new genes and polymorphisms that influence disease risk and have disease-modifying effects will provide a better understanding of the molecular pathogenesis of s-IBM and may lead to new therapeutic targets being identified for treatment of the disease.

Acknowledgements

Data collection and sharing for this project was funded by the Alzheimer's Disease Neuroimaging Initiative (ADNI) (National Institutes of Health Grant U01 AG024904). ADNI is funded by the National Institute on Aging, the National Institute of Biomedical Imaging and Bioengineering, and through generous contributions from the following: Alzheimer's Association; Alzheimer's Drug Discovery Foundation; BioClinica, Inc.; Biogen Idec Inc.; Bristol-Myers Squibb Company; Eisai Inc.; Elan Pharmaceuticals, Inc.; Eli Lilly and Company; F. Hoffmann-La Roche Ltd and its affiliated company Genentech. Inc.: GE Healthcare: Innogenetics. N.V.: IXICO Janssen Alzheimer Ltd.: Immunotherapy Research & Development, LLC.; Johnson & Johnson & Development Pharmaceutical Research LLC.: Medpace, Inc.; Merck & Co., Inc.; Meso Scale Diagnostics, LLC.; Research; NeuroRx Novartis Pharmaceuticals Corporation; Pfizer Inc.; Piramal Imaging: Servier; Synarc Inc.; and Takeda Pharmaceutical Company. The Canadian Institutes of Health Research is providing funds to support ADNI clinical sites in Canada. Private sector contributions are facilitated by the Foundation for the National Institutes of Health (www.fnih.org). The grantee organization is the Northern California Institute for Research and Education, and the study is coordinated by the Alzheimer's Disease Cooperative Study at the University of California, San Diego. ADNI data are disseminated by the Laboratory for Neuro-Imaging at the University of California, Los Angeles. This research was also supported by NIH Grants P30 AG010129 and K01 AG030514.

We thank Dr V. Fabian and Dr. R. Junckerstorff from the Section of Neuropathology, Royal Perth Hospital and the other pathologists who reported the muscle biopsies, and the clinicians who referred patients. Dr A Rojana-udomsart was supported by the Enid and Arthur Home Memorial Scholarship and a University of Western Australia Scholarship for International Research Fees (SIRF). Financial support for the study was provided by Zinfandel and by the Neuromuscular Foundation of Western Australia.

References

- Dalakas MC. Sporadic inclusion body myositis-diagnosis, pathogenesis and therapeutic strategies. Nat Clin Pract Neurol 2006;2:437–47.
- [2] Needham M, Mastaglia FL. Inclusion body myositis: current pathogenetic concepts and diagnostic and therapeutic approaches. Lancet Neurol 2007;6:620–31.

- [3] Needham M, Mastaglia FL. Sporadic inclusion body myositis: a continuing puzzle. Neuromuscul Disord 2008;18:6–16.
- [4] Askanas V, Engel WK, Nogalska A. Pathogenic considerations in sporadic inclusion-body myositis, a degenerative muscle disease associated with aging and abnormalities of myoproteostasis. J Neuropathol Exp Neurol 2012;71:680–93.
- [5] Corder EH, Saunders AM, Strittmatter WJ, et al. Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. Science 1993;261:921–3.
- [6] Needham M, Hooper A, James I, et al. Apolipoprotein epsilon alleles in sporadic inclusion body myositis: a reappraisal. Neuromuscul Disord 2008;18:150–2.
- [7] Askanas V, Engel WK, Mirabella M, et al. Apolipoprotein E alleles in sporadic inclusion-body myositis and hereditary inclusion-body myopathy. Ann Neurol 1996;40:264–5.
- [8] Mirabella M, Alvarez RB, Engel WK, Weisgraber KH, Askanas V. Apolipoprotein E and apolipoprotein E messenger RNA in muscle of inclusion body myositis and myopathies. Ann Neurol 1996;40: 864–72.
- [9] Rojana-udomsart A, James I, Castley A, et al. High-resolution HLA-DRB1 genotyping in an Australian inclusion body myositis (s-IBM) cohort: an analysis of disease-associated alleles and diplotypes. J Neuroimmunol 2012;250:77–82.
- [10] Rojana-Udomsart A, Mitrpant C, James I, et al. Analysis of HLA-DRB3 alleles and supertypical genotypes in the MHC Class II region in sporadic inclusion body myositis. J Neuroimmunol 2012;254: 174–7.
- [11] Mastaglia FL, Needham M, Scott A, et al. Sporadic inclusion body myositis: HLA-DRB1 allele interactions influence disease risk and clinical phenotype. Neuromuscul. Disord. 2009;19:763–5.
- [12] Kok CC, Boyt A, Gaudieri S, et al. Mitochondrial DNA variants in inclusion body myositis. Neuromuscul. Disord. 2000;10:604–11.
- [13] Hansson Petersen CA, Alikhani N, Behbahani H, et al. The amyloid beta-peptide is imported into mitochondria via the TOM import machinery and localized to mitochondrial cristae. Proc Natl Acad Sci USA 2008;105:13145–50.
- [14] Roses AD, Lutz MW, Amrine-Madsen H, et al. A TOMM40 variable-length polymorphism predicts the age of late-onset Alzheimer's disease. Pharmacogenomics J 2010;10:375–84.
- [15] Maruszak A, Peplonska B, Safranow K, Chodakowska-Zebrowska M, Barcikowska M, Zekanowski C. TOMM40 rs10524523 polymorphism's role in late-onset Alzheimer's disease and in longevity. J Alzheimers Dis 2012;28:309–22.
- [16] Cruchaga C, Nowotny P, Kauwe JS, et al. Association and expression analyses with single-nucleotide polymorphisms in TOMM40 in Alzheimer disease. Arch. Neurol. 2011;68:1013–9.
- [17] Crenshaw DG, Gottschalk WK, Lutz MW, et al. Using genetics to enable studies on the prevention of Alzheimer's disease. Clin. Pharmacol. Ther. 2013;93:177–85.
- [18] Griggs RC, Askanas V, DiMauro S, et al. Inclusion body myositis and myopathies. Ann Neurol 1995;38:705–13.
- [19] Linnertz C, Saunders AM, Lutz MW, et al. Characterization of the poly-T variant in the TOMM40 gene in diverse populations. PLoS ONE 2012;7:e30994.
- [20] Oldfors A, Larsson NG, Lindberg C, Holme E. Mitochondrial DNA deletions in inclusion body myositis. Brain 1993;116(Pt 2):325–36.
- [21] Santorelli FM, Sciacco M, Tanji K, et al. Multiple mitochondrial DNA deletions in sporadic inclusion body myositis: a study of 56 patients. Ann Neurol 1996;39:789–95.
- [22] Horvath R, Fu K, Johns T, Genge A, Karpati G, Shoubridge EA. Characterization of the mitochondrial DNA abnormalities in the skeletal muscle of patients with inclusion body myositis. J Neuropathol Exp Neurol 1998;57:396–403.
- [23] Yang CC, Alvarez RB, Engel WK, Askanas V. Increase of nitric oxide synthases and nitrotyrosine in inclusion-body myositis. NeuroReport 1996;8:153–8.
- [24] Schmidt J, Barthel K, Zschuntzsch J, et al. Nitric oxide stress in sporadic inclusion body myositis muscle fibres: inhibition of inducible

nitric oxide synthase prevents interleukin-1beta-induced accumulation of beta-amyloid and cell death. Brain 2012;135:1102–14.

- [25] Askanas V, Engel WK. Sporadic inclusion-body myositis and hereditary inclusion-body myopathies: diseases of oxidative stress and aging? Arch Neurol 1998;55:915–20.
- [26] Hashimoto M, Rockenstein E, Crews L, Masliah E. Role of protein aggregation in mitochondrial dysfunction and neurodegeneration in Alzheimer's and Parkinson's diseases. Neuromolecular Med 2003;4:21–36.
- [27] Askanas V, Engel WK. Inclusion-body myositis: a myodegenerative conformational disorder associated with Abeta, protein misfolding, and proteasome inhibition. Neurology 2006;66:S39–48.
- [28] Askanas V, McFerrin J, Baque S, Alvarez RB, Sarkozi E, Engel WK. Transfer of beta-amyloid precursor protein gene using adenovirus

vector causes mitochondrial abnormalities in cultured normal human muscle. Proc Natl Acad Sci USA 1996;93:1314–9.

- [29] Anandatheerthavarada HK, Biswas G, Robin MA, Avadhani NG. Mitochondrial targeting and a novel transmembrane arrest of Alzheimer's amyloid precursor protein impairs mitochondrial function in neuronal cells. J Cell Biol 2003;161:41–54.
- [30] Devi L, Prabhu BM, Galati DF, Avadhani NG, Anandatheerthavarada HK. Accumulation of amyloid precursor protein in the mitochondrial import channels of human Alzheime's disease brain is associated with mitochondrial dysfunction. J Neurosci 2006;26: 9057–68.
- [31] Crouch PJ, Blake R, Duce JA, et al. Copper-dependent inhibition of human cytochrome c oxidase by a dimeric conformer of amyloidbeta1-42. J Neurosci 2005;25:672–9.